

In-Depth Search Focused on Furans, Lactones, Volatile Phenols, and Acetals As Potential Age Markers of Madeira Wines by Comprehensive Two-Dimensional Gas Chromatography with Time-of-Flight Mass Spectrometry Combined with Solid Phase Microextraction

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ABSTRACT: The establishment of potential age markers of Madeira wine is of paramount significance as it may contribute to detect frauds and to ensure the authenticity of wine. Considering the chemical groups of furans, lactones, volatile phenols, and acetals, 103 volatile compounds were tentatively identified; among these, 71 have been reported for the first time in Madeira wines. The chemical groups that could be used as potential age markers were predominantly acetals, namely, diethoxymethane, 1,1-diethoxyethane, 1,1-diethoxy-2-methyl-propane, 1-(1-ethoxyethoxy)-pentane, *trans*-dioxane and 2-propyl-1,3-dioxolane, and from the other chemical groups, 5-methylfurfural and *cis*-oak-lactone, independently of the variety and the type of wine. GC × GC-ToFMS system offers a more useful approach to identify these compounds compared to previous studies using GC-qMS, due to the orthogonal systems, that reduce coelution, increase peak capacity and mass selectivity, contributing to the establishment of new potential Madeira wine age markers. Remarkable results were also obtained in terms of compound identification based on the organized structure of the peaks of structurally related compounds in the GC × GC peak apex plots. This information represents a valuable approach for future studies, as the ordered-structure principle can considerably help the establishment of the composition of samples. This new approach provides data that can be extended to determine age markers of other types of wines.

KEYWORDS: Age markers, Madeira wine, HS-SPME, GC × GC-ToFMS

INTRODUCTION

Madeira wine is a fortified Portuguese wine produced in Madeira Island over the last centuries and plays an important role in the economy of the Island. The peculiar characteristics of Madeira wines arise from the specific and singular winemaking process. The fermentation process is stopped by the addition of natural grape spirit in order to obtain an ethanol content of 18–22% (v/v). Some wines undergo aging in wood casks in cellars at temperatures up to 30 °C, and humidity levels between 70 and 75%, while the majority of wines are submitted to a baking process, i.e., the wine is placed in large coated vats, and the temperature is slowly increased at about 5 °C per day and maintained at 45–50 °C during at least 3 months. After this treatment, the wine is allowed to undergo a maturation process in oak casks for a minimum of 3 years. Finally, some Madeira wines were submitted to an aging process, from a minimum of 3 to 20 years or even longer.^{1,2} The aging process in oak casks is fundamental for the Madeira wine's unique sensorial properties. During this period, several reactions and migration of molecules from the oak to wine can occur,^{3,4} which depends on some parameters, such as grape variety, wine making procedure, and oak characteristics (geographical origin, species of oak, seasoning of the staves, toasting, and age of cask),^{5–9} among others.

The establishment of potential age markers is important to detect frauds and to ensure the authenticity of the wine. Furthermore, the economic value of Madeira wine is highly associated

with its age. Some volatile compounds that belong to furans, lactones, volatile phenols, and acetals have been reported as potential aging markers in Madeira wines.^{4,10–12} Compounds such as, 2-furfural, 5-methylfurfural, 5-hydroxymethylfurfural, *cis*-oak-lactone, *trans*-oak-lactone, eugenol, guaiacol, *m*-cresol, *o*-cresol, *p*-ethylphenol, maltol, vanillin, *cis*-dioxane, *trans*-dioxane, *cis*-dioxolane, and *trans*-dioxolane were considered.^{5,13–16} Furans (e.g., 2-furfural, 5-methylfurfural, and 5-hydroxymethyl-2-furfural) are formed by three pathways: pyrolysis of carbohydrates, dehydration of sugars through Maillard reaction, and caramelization,^{17–19} which occurs during winemaking and aging. As the levels of 2-furfural and 5-hydroxymethyl-2-furfural have a tendency to increase linearly during aging, they were considered as age markers.^{4,20} The lactones are important flavor compounds which are produced by cyclization of the corresponding hydroxycarboxylic acids.^{4,21} Oak lactones, such as *cis*- and *trans*-oak-lactone, are already present in natural oak, and their content increased due to seasoning and toasting,¹⁴ and from an organoleptic point of view, they are the most important lactones extractable from oak casks.²² Volatile phenols, like ethyl and vinylphenols, were also extracted from oak; nevertheless, their microbiological yeast transformation (e.g., *Brettanomyces* and

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Dekkara) from hydroxycinnamic acids of wine were reported as the main origin.²³ Acetals are formed during fermentation; nevertheless, their content increases significantly during the oxidative conditions of aging process. The high acetaldehyde content in wine contributes to the acetalization reaction with glycerol, which is favored at higher pH values, leading to four heterocyclic acetal alcohol formation: *cis*- and *trans*-5-hydroxy-2-methyl-1,3-dioxane (*cis*-dioxane and *trans*-dioxane), and *cis*- and *trans*-4-hydroxymethyl-2-methyl-1,3-dioxolane (*cis*-dioxolane and *trans*-dioxolane). Heterocyclic acetal alcohols were identified and reported as potential age markers of Madeira wine.^{4,10,11} Other acetals, such as 1,1-diethoxyethane and 2,4,5-trimethyl-dioxolane, were also detected in table wines.²⁴

The Madeira wine volatile composition related to aging process has been studied using a one-dimensional chromatographic (¹D-GC) process, which revealed the complexity of this matrix.^{4,10–12} Although such a method often provides rewarding analytical results, in-depth analysis of the chromatograms frequently indicates that some peaks are the result of two or more coeluting compounds. Comprehensive two-dimensional gas chromatography (GC × GC) was developed as a powerful separation method and emerged as an interesting alternative to analyze complex samples or analyze trace target analytes within a single analysis and overcoming the coelution problem.²⁵ The method employs two orthogonal mechanisms and is based on the application of two GC columns coated with different stationary phases, a nonpolar and a polar one (NP/P), sequentially linked through a modulator. Thus, the separation is ruled by boiling point properties in the first dimension (¹D) and polarity in the second one (²D).^{26,27} Therefore, two-dimensional gas chromatography (GC × GC) offers faster running times, increased peak capacity, improved resolution and enhanced mass selectivity, good calibration linearity, and more sensitivity, and the limits of detection are improved due to the focusing of the peak in the modulator when compared to that in the one-dimensional GC.^{28–30}

In order to obtain a deeper characterization of the chemical groups potentially related with Madeira wine aging, namely, furans, lactones, volatile phenols, and acetals, the comprehensive two-dimensional gas chromatography with time-of-flight mass spectrometry (GC × GC–ToFMS) combined with headspace solid-phase microextraction (HS-SPME) was used in the present research. This methodology was applied to Madeira wines from different varieties (Malvasia, Bual, Sercial, Verdelho, and Tinta Negra), types (sweet, medium sweet, dry, and medium dry), and ages (Vintage and blended wines). Finally, principal component analysis (PCA) was applied in order to establish potential age markers, which allow one to distinguish the different types of Madeira wines based on their age and even blends (average age).

MATERIAL AND METHODS

Samples. Twenty-three monovarietal Madeira wines from five *Vitis vinifera* L. grape varieties (one red, Tinta Negra, and four white, named as noble varieties of Madeira wine, Malvasia, Bual, Sercial, and Verdelho), aged from 3 to 20 years old (Y) and matured in oak casks, were used in this study. Tinta Negra is the main grape variety harvested in Madeira Island (Portugal) representing more than 80% of the vineyards. According to the age, the wines under study correspond to Vintage (a specific year of aged in casks, 17, 18, 19, and 20 years) and blended (B, an average aging period of 3, 5, 10, or 15 years) wines. Four types of wine were used: sweet (Malvasia, Tinta Negra), medium sweet (Bual, Tinta Negra), dry (Sercial, Tinta Negra), and medium dry (Verdelho, Tinta Negra), and

were aged in American oak casks (submitted to a lighter toasting). The ethanol content of the Madeira wines under study ranged from 18 to 19% (v/v). The samples were kindly provided by Madeira Wine Company, Madeira Island.

Reagents and Standards. Sodium chloride (99.5%, foodstuff grade) was purchased from Sigma Aldrich (Madrid, Spain), and ultra pure water was obtained from a Milli-Q system from Millipore (Milford, MA, USA). The retention index probes (*n*-alkanes series of C₈ to C₂₀ straight-chain alkanes, concentration 40 mg/L in *n*-hexane) were supplied from Fluka (Buchs, Switzerland).

HS-SPME Methodology. The HS-SPME experimental parameters were previously established.² The SPME holder for manual sampling and fiber were purchased from Supelco (Aldrich, Bellefonte, PA, USA). The SPME device included a fused silica fiber coating partially cross-linked with 50/30 μm divinylbenzene-carboxen-poly(dimethylsiloxane). Prior to use, the SPME fiber was conditioned at 270 °C for 60 min in the GC injector, according to the manufacturer's recommendations. Then, the fiber was daily conditioned for 10 min at 250 °C.

For the HS-SPME assay, aliquots of 1 mL of the sample were placed into a 5 mL glass vial. After the addition of 0.5 g of NaCl and stirring (0.5 × 0.1 mm bar) at 400 rpm, the vial was capped with a PTFE septum and an aluminum cap (Chromacol, Hertfordshire, UK). The vial was placed in a thermostatted bath adjusted to 60.0 ± 0.1 °C for 5 min, and then the SPME fiber was inserted in the headspace for 20 min. Each sample was analyzed in triplicate. Blanks, corresponding to the analysis of the coating fiber not submitted to any extraction procedure, were run between sets of three analyses.

GC × GC–ToFMS Analysis. The GC × GC–ToFMS methodology was based on a previous study.² After the extraction/concentration step, the SPME coating fiber was manually introduced into the GC × GC–ToFMS injection port at 250 °C and kept for 3 min for desorption. The injection port was lined with a 0.75 mm I.D. splitless glass linear. Splitless injections were used (30 s). LECO Pegasus 4D (LECO, St. Joseph, MI, USA) GC × GC–ToFMS system consisted of an Agilent GC 7890A gas chromatograph, with a dual stage jet cryogenic modulator (licensed from Zoex) and a secondary oven. The detector was a high-speed ToF mass spectrometer. An HP-5 column (30 m × 0.32 mm I.D., 0.25 μm film thickness, J&W Scientific Inc., Folsom, CA, USA) was used as first-dimension column, and a DB-FFAP (0.79 m × 0.25 mm I.D., 0.25 μm film thickness, J&W Scientific Inc., Folsom, CA, USA) was used as a second-dimension column. The carrier gas was helium at a constant flow rate of 2.50 mL/min. The primary oven temperature was programmed from 40 (1 min) to 230 °C (2 min) at 10 °C/min. The secondary oven temperature was programmed from 70 (1 min) to 250 °C (3 min) at 10 °C/min. The MS transfer line temperature was 250 °C, and the MS source temperature was 250 °C. The modulation time was 6 s; the modulator temperature was kept at 20 °C offset (above primary oven). A 6 s modulation time with a 30 °C secondary oven temperature offset was chosen to be a suitable compromise as it maintained the 1D separation, maximized the 2D resolution, and avoided the wrap-around effect (the elution time of a pulsed solute exceeds the modulation period) for compounds that were late to elute from the 2D. Ideally, all peaks must be detected before the subsequent reinjection, and hence, ²t_R must be equal or less than the modulation period.^{31,32} The ToFMS was operated at a spectrum storage rate of 125 spectra/s. The mass spectrometer was operated in the EI mode at 70 eV using a range of *m/z* 33–350, and the voltage was –1695 V. Total ion chromatograms (TIC) were processed using the automated data processing software ChromaTOF (LECO) at a signal-to-noise threshold of 10. Contour plots were used to evaluate the separation general quality and for manual peak identification; a signal-to-noise threshold of 50 was used. Two commercial databases (Wiley 275 and US National Institute of Science and Technology (NIST) V. 2.0, Mainlib and Replib) were used. A mass spectral match factor, the majority (86%) of the tentatively

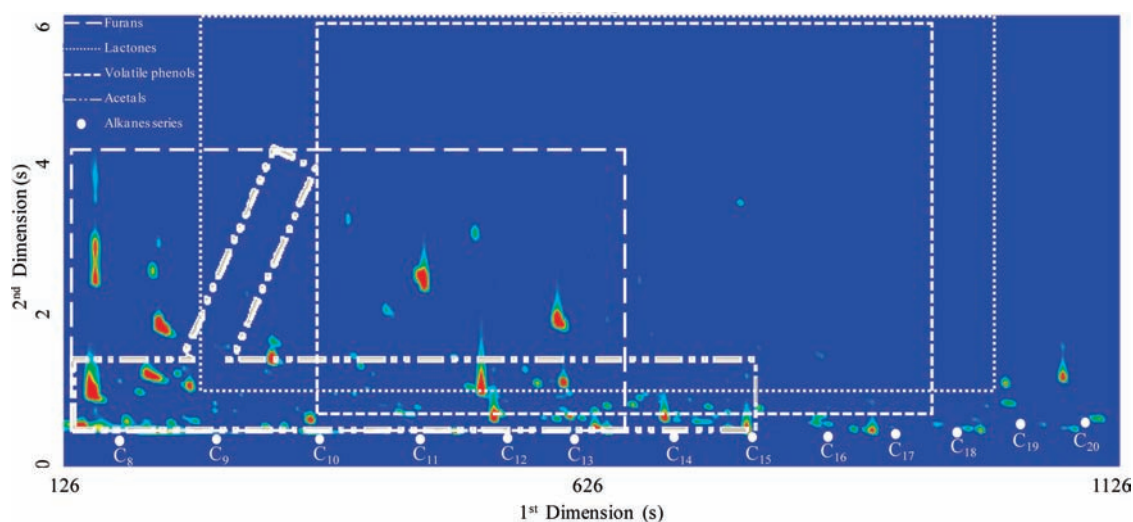


Figure 1. GC \times GC total ion current chromatogram contour plot obtained from a sweet Madeira wine (Tinta Negra, 5 years); the chromatographic spaces corresponding to furans, lactones, volatile phenols, and acetals were highlighted. The *n*-alkanes series (C_6 – C_{20}) was superimposed on the contour plot.

identified compounds showed similarity matches >850 , was set to decide whether a peak was correctly identified or not. Furthermore, a manual inspection of the mass spectra was done, combined with the use of additional data, such as the retention index (RI) value, which was determined according to the Van den Dool and Kratz RI equation.³³ For the determination of the RI, a C_8 – C_{20} *n*-alkanes series was used, and as some volatile compounds were eluted before C_8 , the solvent *n*-hexane was used as the C_6 standard. The RI values experimentally calculated were compared, when available, with values reported in the literature for similar chromatographic columns that employ as the first dimension the column of this study.^{34–51} The GC \times GC area data were used as an approach to estimate the relative content of each volatile component.

Data Processing. In an initial approach, a linear regression was performed between total GC peak area of the chemical groups (furans, lactones, volatile phenols, and acetals) under study and wine age in order to establish potential age markers for Malvasia and Bual wines, and the results were expressed as r^2 (coefficient of determination). In a second step, PCA (principal component analysis) was applied to the autoscaled areas of the 103 volatile compounds tentatively identified by HS-SPME/GC \times GC–ToFMS present in 23 monovarietal Madeira wines (from different varieties, types, and age) each with three replicates, using the R statistical software package.⁵² Autoscaling is a data pretreatment process that makes variables of different scales comparable. Each variable is autoscaled separately by subtracting its mean value and dividing by its standard deviation. The goal was to extract the main sources of variability and hence to help with the characterization of the data set.⁵³

RESULTS AND DISCUSSION

Contour and Peak Apex Plot Analysis. Automated processing of HS-SPME/GC \times GC–ToFMS data was used to tentatively identify all peaks in the GC \times GC chromatogram contour plots with a signal-to-noise threshold >50 . The contour plot of the total ion chromatogram (Figure 1) exhibited several hundreds of peaks; however, this study was only focused on furans, lactones, volatile phenols, and acetals. The peak finding routine based on the deconvolution method allowed us to detect 103 compounds from these four chemical groups, which were tentatively identified on the basis of the comparison of their mass spectra to a reference database (MS) and by comparison of the RIs calculated (RI_{calc}) with the values reported in the literature (RI_{lit})

for the 5% phenylpolysilphenylene-siloxane (or equivalent) column (Tables 1 and 2). A range between 1 and 30 ($|RI_{\text{calc}} - RI_{\text{lit}}|$) was obtained for RI_{calc} compared to the RI_{lit} reported in the literature for one-dimensional GC with the 5%-phenyl-methylpolysiloxane GC column or equivalent. This difference in RI is considered reasonable ($<5\%$) if one takes into account that (i) the literature data is obtained from a large range of GC stationary phases (several commercial GC columns are composed of 5% phenylpolysilphenylene-siloxane or equivalent stationary phases) and that (ii) the literature values were determined in a one-dimensional chromatographic separation system, and the modulation causes some inaccuracy in the first dimension retention time.⁵⁴ In the case of the volatile compounds with $|RI_{\text{calc}} - RI_{\text{lit}}|$ values higher than 30, the information related to the mass spectra (m/z) was included in Tables 1 and 2.

Figure 1 shows the GC \times GC total ion current chromatogram contour plot obtained from a sweet Madeira wine (Tinta Negra, 5Y); the chromatographic spaces corresponding to furans, lactones, volatile phenols, and acetals were highlighted. The *n*-alkanes series (C_6 – C_{20}) used for the calculation of experimental RIs are also superimposed on the contour plot. The components of each chemical group were dispersed through the contour plot according to their volatility (1^{D}) and polarity (2^{D}), and it becomes difficult to establish the two-dimension chromatographic space (GC \times GC) specific for each chemical group. As the principle of the structured chromatogram is very important in the identification, especially for the compounds that are not commercially available, a strategy was implemented to find this principle. Thus, peak apex plots were constructed, in order to find the possible structured 2D chromatographic profile, combining 1^{D} and 2^{D} values, for each chemical group under study, as shown for furans (Figure 2), lactones (Figure 3), volatile phenols (Figure 4), and acetals (Figure 5). Peak apex plots indicate the position of the maximum modulated peak of GC \times GC analysis, in the 2D chromatographic space.⁵⁵ For all chemical groups, as expected, it was observed that the decrease in volatility (high 1^{D}) is mainly related to the increase in the number of carbons.

The furans include several types of chemical structures; thus, they were organized in furan/alkyl furan, furanic aldehyde, furanic alcohol, benzofuran, furanic ester, and furanic acetal

Table 1. Continued

Peak number	$^1f_R^a$ (s)	$^2f_R^b$ (s)	Chemical groups	Compounds previously identified in Madeira wines	R.I. Calc. ^b	R.I. Lit. ^c	R.I. Lit. ^d	dry						medium dry												
								Sercial			Tinta Negra			Verdelho			Tinta Negra									
								10Y B	3Y B	SY B	10Y B	3Y B	SY B	10Y B	3Y B	SY B	10Y B	3Y B	SY B							
13	240	490	Lactones	-	881	828	-	0.68	(8)	1.97	(7)	0.44	(28)	0.73	(8)	0.17	(16)	0.47	(23)	5.26	(5)	-	-	-		
20	282	253	2,5-Furandione ($m/z = 98, 54, 26$) ^f	(4, 11, 12, 60)	932	915	920	57.68	(19)	37.92	(1)	36.78	(8)	60.06	(13)	41.87	(16)	38.52	(13)	52.76	(12)	30.54	(14)	58.97	(19)	
21	282	4.15	γ -Butyrolactone	-	933	915	-	5.11	(11)	3.76	(3)	2.29	(15)	2.52	(15)	3.99	(16)	4.14	(10)	2.77	(8)	3.15	(12)	2.90	(7)	
25	306	2.67	γ -Crotonolactone	-	956	920	-	0.19	(8)	0.29	(1)	0.81	(19)	0.21	(5)	0.47	(12)	0.46	(11)	0.48	(13)	0.58	(7)	0.85	(4)	
26	306	2.99	α -Angelicallactone ($m/z = 98, 55, 43$) ^f	-	956	-	-	5.11	(32)	5.85	(29)	0.84	(21)	5.92	(14)	8.62	(7)	11.30	(8)	3.85	(11)	1.66	(12)	-	-	
27	318	1.98	3-Methylenehydro-2,5-furandione ($m/z = 112, 84, 68, 40$) ^f	-	967	943	-	0.93	(13)	0.91	(3)	0.73	(10)	0.49	(18)	0.69	(9)	0.81	(10)	1.03	(16)	0.38	(11)	0.30	(7)	
34	342	2.55	γ -Pentalactone	-	991	979	-	1.73	(12)	4.02	(8)	1.80	(20)	1.97	(12)	2.34	(4)	2.05	(5)	5.58	(16)	4.64	(10)	3.82	(5)	
36	348	2.99	α -Methyl- γ -crotonolactone	-	997	967	-	0.20	(28)	0.21	(5)	0.14	(3)	0.15	(17)	0.21	(27)	0.28	(12)	0.23	(6)	0.23	(13)	0.39	(8)	
41	360	1.52	2H-Pyran-2-one	-	1008	992	-	0.19	(6)	-	-	-	-	-	-	-	-	-	-	-	-	0.25	(8)	0.39	(15)	
44	384	4.38	$\beta\beta$ -Dimethylbutyrolactone	-	1036	1057	-	1.73	(8)	4.98	(8)	0.66	(8)	4.76	(23)	4.23	(11)	4.30	(6)	6.14	(25)	2.15	(3)	2.12	(15)	
47	396	1.66	3,4-Dihydro-3-methyl-2,5-furandione	-	1046	1039	1045	0.30	(3)	0.32	(7)	-	-	-	-	-	-	-	-	-	-	0.32	(9)	0.30	(5)	
48	396	2.02	Lavander lactone	-	1046	-	-	1.29	(14)	1.16	(5)	0.56	(19)	0.67	(15)	0.99	(2)	1.18	(15)	1.12	(13)	0.63	(5)	4.29	(22)	
49	396	5.90	3,4-Dimethyl-2,5-furandione ($m/z = 126, 82, 54, 39$) ^f	-	1050	-	-	1060	2.54	(15)	5.81	(9)	1.33	(35)	4.34	(13)	4.19	(7)	3.01	(12)	3.93	(9)	1.21	(12)	2.31	(6)
50	402	3.30	Pantolactone ($m/z = 131, 71, 57, 43$) ^f	(4, 12)	1053	1036	-	0.37	(3)	0.79	(6)	0.35	(15)	0.28	(29)	0.34	(20)	0.35	(18)	0.34	(19)	0.50	(14)	1.71	(12)	
52	408	1.76	4-Methyl-2-(SH)-furanone	-	1058	1055	1060	1.20	(14)	0.95	(14)	0.44	(10)	0.83	(13)	0.77	(11)	0.64	(11)	0.75	(17)	1.70	(26)	2.66	(11)	
53	408	2.43	γ -Hexalactone	(4)	1059	-	-	-	0.60	(11)	4.13	(22)	3.14	(22)	3.14	(6)	4.76	(14)	3.09	(34)	1.69	(8)	3.18	(18)	5.99	(14)
55	420	1.75	β -methyl- γ -butyrolactone ($m/z = 85, 56, 41$) ^f	-	1071	1067	1069	5.89	(3)	20.33	(11)	4.36	(12)	13.41	(19)	11.15	(13)	1.43	(4)	11.53	(15)	3.64	(8)	5.86	(2)	
71	480	3.87	γ -Ethoxybutyrolactone	-	1140	1107	-	0.38	(8)	1.15	(8)	0.34	(8)	0.32	6	0.76	(9)	0.88	(2)	0.90	(16)	0.38	(8)	0.43	(7)	
72	492	2.43	Solerone ($m/z = 118, 56, 41$) ^f	(4)	1152	1130	1162	0.44	(18)	0.81	(9)	-	-	-	-	0.84	(16)	0.47	(8)	0.78	(6)	-	-	-	-	
73	510	2.86	γ -Heptalactone	-	1174	1156	-	1.22	(4)	2.29	(6)	0.40	(20)	1.18	(14)	1.84	(15)	0.95	(10)	1.88	(11)	0.38	(16)	1.04	(5)	
80	588	1.50	Mevalonic lactone	-	1284	1287	1291	0.63	(6)	0.75	(7)	0.44	(10)	0.46	(5)	0.80	(18)	0.55	(8)	0.59	(9)	0.67	(11)	1.30	(13)	
83	618	1.26	γ -Octalactone	(4, 60)	1320	1292	1297	7.72	(2)	14.80	(10)	0.50	(12)	1.33	(9)	7.81	(8)	9.17	(10)	24.72	(16)	1.62	(19)	2.02	(15)	
88	642	1.37	<i>trans</i> -oak-lactone	(4, 11, 60)	1345	1340	1329	24.84	(6)	58.99	(7)	2.37	(8)	4.85	(10)	28.14	(4)	32.03	(9)	52.87	(22)	4.94	(11)	6.54	(13)	
89	660	3.39	<i>cis</i> -oak-lactone	-	1366	-	-	0.29	(15)	0.41	(19)	-	-	0.13	(19)	0.36	(8)	0.38	(15)	0.31	(25)	0.29	(15)	0.63	(16)	
91	672	1.44	2-Benzofuran-1(3H)-one ($m/z = 134, 105, 77, 51$) ^f	-	1377	1366	1368	2.25	(11)	2.61	(12)	1.66	(18)	1.15	(11)	3.83	(6)	2.13	(5)	1.39	(10)	2.30	(17)	3.98	(2)	
96	750	1.40	γ -Nonalactone	(4, 60)	1471	1472	1472	0.69	(6)	0.61	(8)	0.64	(18)	-	-	0.87	(4)	0.47	(18)	0.24	(13)	0.74	(13)	1.30	(17)	
97	762	1.64	γ -Decalactone	-	1487	1508	1482	0.37	(9)	0.47	(11)	0.12	(27)	0.10	(26)	0.33	(9)	0.23	(6)	0.15	(12)	0.25	(17)	0.33	(18)	
100	900	1.25	Massoia lactone	-	1684	1685	-	0.43	(19)	0.27	(9)	0.21	(16)	0.09	(1)	0.57	(18)	0.13	(7)	0.07	(13)	0.20	(15)	1.24	(11)	
101	918	1.33	γ -Dodecalactone	-	1713	1721	-	0.30	(23)	0.20	(22)	0.15	(15)	-	-	0.14	(24)	0.17	(11)	-	-	-	0.28	(6)	0.84	(10)
103	996	0.87	δ -Dodecalactone	-	1841	1839	-	2.54	(12)	1.15	(9)	-	-	-	-	3.21	(11)	1.20	(12)	-	-	-	-	4.32	(8)	
			Muskolactone	-	1722	1722	1722	8.17	(8)	174.39	(5)	62.49	(4)	109.10	(11)	134.27	(6)	120.81	(3)	181.36	(9)	66.83	(8)	116.81	(10)	
			Subtotal	-																						

Table 1. Continued

Peak number	$t_{R'}^a$ (s)	$^2 t_{R'}^a$ (s)	Chemical groups	Compounds previously identified in Madeira wines	R.I. Calc. ^b	R.I. Lit. ^c	R.I. Lit. ^d	dry						medium dry												
								Sercial			Tinta Negra			Verdelho			Tinta Negra									
								10Y B	3Y B	SY B	10Y B	3Y B	SY B	10Y B	3Y B	SY B	10Y B	3Y B	SY B							
Volatile Phenols																										
39	354	1.74	Phenol	(59)	1002	980	996	0.64	(7)	7.33	(3)	-	-	4.52	(8)	1.37	(4)	8.48	(9)	4.98	(13)	3.78	(17)	10.27	(33)	
43	378	0.94	<i>o</i> -Methylanisole	-	1026	1001	-	0.17	(11)	0.37	(7)	-	-	-	-	0.27	(13)	0.25	(4)	0.61	(8)	-	-	0.33	(24)	
54	414	4.86	<i>o</i> -Cresol	(60)	1068	1053	1069	1.87	(4)	1.56	(2)	1.42	(10)	1.99	(2)	1.93	(11)	1.44	(10)	0.96	(6)	1.79	(14)	2.99	(18)	
58	432	5.78	<i>p</i> -Cresol	-	1087	1072	1091	1.44	(9)	1.12	(14)	-	-	1.02	(16)	1.29	(11)	0.52	(18)	0.67	(14)	0.82	(6)	1.88	(7)	
62	438	2.47	<i>o</i> -Guaiacol	(60)	1090	1091	1092	2.16	(11)	2.26	(8)	2.17	(11)	2.77	(10)	2.27	(4)	1.46	(7)	2.12	(8)	1.91	(3)	3.11	(13)	
68	462	0.92	<i>p</i> -Ethylanisole	-	1115	1122	-	0.37	(16)	1.17	(5)	0.18	(16)	-	-	0.75	(5)	0.62	(10)	0.57	(9)	0.20	(8)	0.41	(5)	
74	516	4.62	<i>p</i> -Ethylphenol	(59, 60)	1183	1178	1186	4.45	(5)	22.24	(9)	0.91	(8)	1.71	(12)	4.39	(11)	7.39	(7)	7.28	(11)	0.25	(4)	2.27	(19)	
76	534	2.06	<i>p</i> -Methylguaiacol	-	1202	1191	-	0.60	(7)	0.59	(10)	0.51	(3)	0.39	(13)	0.58	(6)	0.31	(7)	0.43	(6)	0.44	(13)	0.65	(5)	
77	558	3.18	2-Phenoxyethanol	(4, 11)	1240	1220	-	1.82	(12)	0.58	(9)	0.31	(6)	0.51	(3)	1.96	(15)	0.46	(8)	0.27	(10)	0.77	(11)	3.79	(13)	
81	606	1.81	<i>p</i> -Ethylguaiacol	(60)	1308	1287	1281	3.24	(6)	12.07	(10)	0.66	(4)	1.05	(5)	3.47	(14)	7.77	(9)	1.77	(7)	1.22	(12)	1.30	(3)	
87	636	2.50	<i>p</i> -Vinylguaiacol	(11, 60)	1340	1323	1319	0.32	(14)	0.95	(9)	0.61	(12)	0.33	(2)	0.30	(12)	0.35	(12)	-	-	-	-	0.26	(17)	
90	666	1.98	Eugenol	(59, 60)	1371	1364	1360	1.32	(5)	1.17	(10)	1.15	(5)	0.69	(18)	1.19	(10)	0.84	(18)	0.92	(17)	1.02	(15)	2.41	(9)	
92	678	1.64	<i>p</i> -Propylguaiacol	(60)	1383	1371	-	0.29	(6)	0.55	(5)	-	-	-	-	0.17	(20)	0.23	(18)	0.22	(4)	0.09	(11)	0.26	(34)	
94	702	1.21	Methylugenol	-	1409	1407	-	3.14	(10)	19.14	(7)	0.36	(7)	0.51	(10)	1.43	(14)	1.32	(10)	4.40	(9)	0.67	(10)	3.40	(2)	
95	708	4.71	Vanillin	(11, 59, 60)	1419	1410	-	0.93	(6)	1.84	(4)	-	-	-	-	1.05	(12)	1.11	(12)	0.59	(10)	0.50	(16)	0.74	(2)	
99	840	2.74	Ethyl vanillate	(11, 12, 60)	1596	1574	-	0.26	(13)	0.44	(24)	0.20	(33)	0.12	(4)	0.35	(13)	0.23	(17)	0.33	(7)	0.61	(21)	0.71	(7)	
102	942	2.15	Nonylphenol	-	1754	1745	-	1.34	(15)	0.92	(13)	1.08	(2)	1.32	(5)	1.07	(15)	0.49	(12)	0.31	(19)	0.76	(13)	2.70	(8)	
			Subtotal						24.35	(2)	74.32	(7)	9.56	(4)	16.92	(4)	23.85	(8)	33.28	(6)	26.41	(8)	14.82	(7)	37.50	(7)
Acetals																										
3	108	0.60	1-Ethoxy-1-methoxyethane	-	634	606	-	-	-	3.04	(18)	-	-	0.25	(16)	-	-	-	-	-	-	-	-	-	-	
4	114	0.50	Diethoxymethane	-	645	670	-	39.88	(14)	35.82	(9)	14.48	(16)	19.57	(12)	33.12	(13)	38.65	(9)	55.78	(8)	18.62	(5)	16.29	(8)	
5	138	0.61	2,4,5-Trimethyl-1,3-dioxolane	-	690	710	-	13.98	(10)	84.93	(12)	45.83	(9)	51.11	(6)	63.66	(12)	137.35	(19)	55.43	(6)	71.32	(6)	82.70	(12)	
6	144	0.54	1,1-Diethoxyethane	-	690	705	-	204.81	(18)	482.10	(13)	146.99	(14)	226.37	(12)	263.81	(18)	271.14	(19)	366.11	(2)	284.80	(19)	242.88	(9)	
7	150	0.68	2-Methyl-1,3-dioxane	-	712	759	-	19.71	(25)	25.82	(11)	5.26	(24)	7.37	(19)	6.05	(10)	4.97	(4)	31.43	(7)	12.21	(8)	8.32	(13)	
			(<i>m/z</i> = 101, 87, 59) ⁱ																							
11	228	0.51	1,1-Diethoxy-2-methyl-propane	-	857	865	859	25.46	(6)	38.60	(2)	3.41	(32)	20.91	(6)	26.75	(2)	38.22	(6)	25.17	(7)	9.11	(4)	14.62	(6)	
14	252	1.77	<i>cis</i> -Dioxane (<i>m/z</i> = 117, 103, 57, 43) ⁱ	(4, 10, 11)	902	-	-	58.99	(10)	55.69	(8)	19.56	(5)	20.13	(18)	46.57	(9)	45.33	(19)	41.07	(8)	15.36	(6)	34.23	(15)	
15	270	0.54	1,1-Diethoxybutane	-	919	904	-	1.13	(4)	0.90	(14)	0.37	(26)	0.70	(1)	0.66	(14)	0.29	(14)	0.85	(20)	0.54	(20)	0.71	(10)	
16	270	2.46	<i>cis</i> -Dioxolane (<i>m/z</i> = 117, 103, 57, 43) ⁱ	(4, 10)	920	-	-	19.89	(9)	16.07	(3)	3.73	(10)	9.30	(13)	14.76	(15)	7.09	(5)	10.22	(18)	4.21	(11)	9.53	(21)	
18	276	0.58	2,6-Diethyl-5-methyl-1,3-dioxan-4-yl acetate	-	924	-	-	12.97	(9)	6.52	(12)	5.98	(9)	3.99	(5)	5.47	(5)	3.45	(9)	8.11	(22)	5.88	(26)	8.23	(2)	
			(<i>m/z</i> = 141, 99, 55, 43) ⁱ																							
19	276	0.63	2-Ethoxytetrahydro-2H-pyran	-	924	945	-	5.43	(6)	3.94	(15)	1.72	(16)	11.99	(15)	4.56	(14)	2.09	(11)	3.91	(5)	4.43	(25)	4.59	(16)	

Table 1. Continued

Peak number	t_{R}^a (s)	${}^2t_{R}^a$ (s)	Chemical groups	Compounds previously identified in Madeira wines	R.I. Calc. ^b	R.I. Lit. ^b	R.I. Lit. ^c	dry						medium dry											
								Sercial			Tinta Negra			Verdelho			Tinta Negra								
								10YB	3YB	5YB	10YB	3YB	5YB	10YB	3YB	5YB	10Y ^f	3YB	5YB						
22	288	2.89	<i>trans</i> -Dioxolane ($m/z = 117, 103, 57, 43$) ^f	(4, 10, 11)	937	-	-	12.53	(13)	12.61	(5)	4.69	(8)	6.38	(15)	11.37	(12)	7.25	(8)	11.31	(17)	3.20	(11)	5.64	(19)
23	294	0.57	2-Ethoxytetrahydro-4-methyl-2H-pyran ($m/z = 129, 85, 55, 43$) ^f	-	942	-	-	1.77	(8)	4.30	(5)	0.16	(16)	0.25	(25)	1.67	(7)	1.34	(20)	1.79	(15)	6.39	(22)	5.25	(12)
24	294	1.26	2,2-Diethoxyethanol	-	942	948	-	1.59	(7)	1.43	(7)	0.93	(12)	1.10	(3)	2.14	(6)	0.38	(18)	1.57	(15)	0.95	(6)	1.86	(17)
28	318	0.53	1,1-Diethoxy-3-methylbutane	-	965	959	953	64.92	(16)	74.47	(2)	18.01	(9)	44.79	(3)	50.60	(5)	36.24	(5)	54.00	(7)	24.57	(7)	35.63	(8)
30	324	0.82	2-Ethyl-5-methyl-1,3-dioxane ($m/z = 129, 101, 55, 41$) ^f	-	971	919	-	0.56	(18)	0.45	(5)	-	-	0.30	(4)	-	-	0.38	(7)	0.49	(12)	-	-	-	-
31	330	0.54	1-(1-Ethoxyethoxy)-pentane	-	977	1004	970	65.55	(10)	67.64	(6)	23.00	(10)	43.67	(15)	52.02	(9)	45.75	(9)	67.52	(7)	30.05	(9)	35.22	(8)
32	330	0.65	4,5-Dimethyl-1,3-dioxane-5-methanol ($m/z = 113, 101, 71, 55, 43$) ^f	-	976	-	-	-	-	-	-	-	-	-	-	-	-	-	-	5.66	(4)	-	-	-	
33	330	4.09	<i>trans</i> -Dioxane ($m/z = 117, 103, 57, 43$) ^f	(4, 10, 11)	980	-	-	38.84	(16)	53.57	(6)	10.82	(10)	27.07	(11)	41.16	(20)	40.62	(15)	47.96	(5)	12.01	(11)	19.09	(4)
40	354	0.55	1,1-Diethoxypropane	-	1001	1004	995	2.26	(17)	2.43	(19)	0.95	(8)	1.34	(9)	1.53	(18)	1.19	(7)	1.73	(19)	1.09	(23)	1.07	(17)
46	390	0.69	2-Propyl-1,3-dioxolane ($m/z = 115, 73, 71, 45$) ^f	-	1038	-	-	8.88	(3)	6.40	(18)	2.47	(2)	2.91	(5)	7.46	(11)	7.47	(1)	7.35	(15)	3.31	(10)	5.56	(19)
59	432	0.62	1,3,3-Triethoxypropane	-	1082	1076	-	6.30	(6)	5.11	(9)	0.94	(17)	1.51	(15)	3.18	(10)	2.71	(8)	3.62	(13)	1.11	(19)	2.24	(7)
64	444	0.56	1,1-Diethoxyhexane	-	1094	1092	1092	8.53	(7)	7.45	(5)	2.25	(8)	4.83	(13)	6.39	(12)	4.71	(5)	5.27	(15)	7.22	(9)	9.06	(8)
66	456	0.57	1-(1-Ethoxyethoxy)-hexane	-	1108	1103	-	4.20	(22)	1.19	(4)	0.76	(21)	2.05	(14)	1.53	(14)	0.98	(10)	1.69	(2)	1.36	(13)	1.16	(10)
69	462	0.62	<i>cis</i> -1,1-Diethoxy-3-hexene	-	1115	1111	-	1.60	(11)	2.24	(5)	0.44	(18)	0.34	(6)	1.57	(9)	1.24	(6)	0.93	(3)	0.90	(10)	1.24	(18)
75	528	0.58	1,1-Diethoxyheptane	-	1192	1202	-	1.27	(5)	1.52	(4)	0.38	(11)	1.11	(14)	0.91	(14)	0.59	(7)	1.02	(9)	0.88	(5)	1.11	(14)
82	612	0.58	1,1-Diethoxyoctane	-	1313	1302	-	1.43	(13)	1.61	(1)	0.99	(6)	2.18	(13)	1.29	(16)	0.89	(3)	0.61	(17)	1.23	(17)	1.58	(8)
84	618	1.07	4,5-Dimethyl-2-phenyl-1,3-dioxolane ($m/z = 196, 152, 77, 43$) ^f	-	1320	-	-	1.47	(1)	2.63	(6)	0.41	(11)	0.77	(6)	0.91	(12)	4.44	(17)	1.21	(15)	0.44	(4)	0.50	(7)
93	684	0.59	1,1-Diethoxynonane	-	1388	1401	1381	4.79	(16)	7.72	(14)	2.80	(12)	5.18	(10)	5.42	(11)	1.96	(19)	2.28	(16)	3.45	(19)	7.16	(2)
98	762	0.60	1,1-Diethoxydecane	-	1485	1501	-	2.17	(12)	5.25	(14)	1.85	(18)	3.99	(11)	2.20	(16)	0.93	(11)	0.97	(7)	4.69	(5)	4.56	(11)
			Subtotal																						

^a Retention times in seconds (s) for first (${}^1t_{R}$) and second (${}^2t_{R}$) dimensions. ^b RI: retention index obtained through the modulated chromatogram. ^c RI: retention index reported in the literature for one dimensional GC with a 5%-Phenyl-methylpolysiloxane GC column or equivalent. ^d RI: retention index reported in the literature for a comprehensive GC × GC system with Equity-5 for the first dimension. ^e Y: vintage wines; age expressed in years. ^f Y: blend wine; years, average age. ^g Mean of three replicates. ^h Relative standard deviation, expressed in percentage. ⁱ Tentatively identified based on mass spectra.

Table 2. Continued

Peak number ¹ _k (s) ² _k (s)	R.I. R.I. R.I. Calc. ^b Lit. ^c Lit. ^d	Chemical groups	sweet										medium sweet																	
			Malvasia					Tinta Negra					Bual					Tinta Negra												
			10Y B	15Y B	18Y ^d	20Y	3Y B	5Y B	10Y B	15Y B	17Y	19Y	3Y B	5Y B	10Y B	15Y B	17Y	19Y	3Y B	5Y B										
13	881 828	-	-	1.36	(9)	1.58	(7)	1.10	(7)	1.03	(8)	-	-	-	2.59	(3)	1.45	(25)	1.00	(26)	0.83	(16)	2.01	(11)	-	-	-			
20	932 915	920 28.61	(18)	27.93	(6)	33.22	(7)	40.74	(9)	58.95	(6)	28.04	(16)	27.34	(2)	27.81	(12)	23.72	(9)	55.78	(10)	88.52	(13)	29.00	(8)	18.53	(4)	35.00	(9)	
21	933 915	-	7.37	(1)	6.36	(10)	15.37	(16)	3.62	(12)	5.62	(13)	3.90	(18)	4.53	(12)	4.37	(6)	2.63	(15)	3.71	(9)	4.99	(13)	7.14	(4)	2.80	(7)	3.78	(10)
25	956 920	-	1.12	(20)	1.08	(2)	1.87	(6)	0.99	(8)	1.34	(10)	0.96	(7)	0.83	(7)	1.60	(22)	1.22	(7)	0.91	(4)	0.52	(5)	0.75	(10)	0.80	(13)	1.17	(12)
26	956	-	-	1.09	(16)	3.73	(2)	3.67	(19)	1.45	(12)	1.52	(22)	0.24	(17)	2.44	(9)	4.71	(18)	4.40	(4)	6.47	(15)	6.35	(8)	6.73	(5)	1.15	(22)	-
27	967 943	-	1.83	(13)	1.11	(15)	0.60	(9)	1.55	(9)	2.19	(3)	0.94	(13)	0.51	(10)	0.64	(11)	0.73	(14)	1.77	(7)	2.74	(10)	1.11	(5)	0.32	(14)	0.73	(13)
34	991 979	-	0.57	(14)	0.56	(11)	4.64	(8)	8.52	(9)	18.43	(5)	4.82	(14)	2.50	(8)	3.91	(19)	10.27	(4)	11.06	(13)	22.63	(17)	1.14	(15)	1.00	(3)	2.84	(18)
36	997 967	-	0.22	(26)	0.25	(13)	0.41	(12)	0.32	(16)	0.40	(10)	-	-	0.66	(17)	0.22	(3)	0.20	(13)	0.19	(15)	0.33	(17)	0.37	(9)	0.17	(13)	-	
41	1008 992	-	-	0.22	(28)	-	-	0.20	(14)	0.35	(20)	-	-	-	-	0.19	(31)	-	-	-	-	-	0.42	(19)	-	-	-	-	-	
44	1036 1057	-	0.26	(18)	0.35	(4)	4.73	(11)	2.06	(19)	2.29	(16)	1.29	(4)	3.18	(13)	4.71	(7)	2.74	(6)	2.63	(8)	2.22	(19)	2.09	(9)	0.28	(13)	1.82	(4)
47	1046 1039	1045.031	(11)	0.41	(14)	0.33	(18)	0.56	(18)	0.33	(33)	0.35	(13)	0.24	(8)	-	-	-	-	0.42	(17)	0.47	(18)	1.08	(2)	-	-	-	-	
48	1046	-	0.67	(11)	1.51	(3)	1.38	(12)	2.43	(10)	1.27	(9)	1.13	(16)	0.68	(8)	0.95	(13)	1.05	(5)	1.42	(19)	1.97	(10)	2.13	(16)	0.38	(7)	0.99	(17)
49	1050	-	10600.87	(10)	1.02	(4)	3.08	(23)	3.09	(3)	2.88	(8)	2.25	(9)	2.63	(17)	1.10	(24)	1.26	(9)	2.53	(14)	4.28	(16)	2.81	(8)	-	-	1.65	(12)
50	1053 1036	-	-	0.60	(12)	1.09	(16)	0.52	(1)	0.70	(24)	0.40	(18)	-	-	0.50	(17)	0.40	(8)	0.53	(6)	0.52	(11)	1.27	(32)	-	-	-	-	
52	1058 1055	10600.60	(6)	0.74	(3)	0.83	(7)	0.75	(5)	1.04	(1)	0.89	(16)	0.47	(4)	0.89	(20)	0.65	(13)	1.03	(11)	1.24	(9)	0.79	(13)	0.36	(9)	0.97	(11)	
53	1059	-	2.74	(5)	3.74	(2)	3.51	(9)	3.29	(19)	4.12	(11)	2.83	(4)	2.91	(8)	0.80	(11)	2.45	(15)	3.00	(19)	2.83	(11)	4.24	(16)	-	-	3.08	(5)
55	1071 1067	10690.56	(30)	12.15	(19)	12.51	(11)	12.22	(11)	15.19	(8)	5.92	(14)	5.91	(11)	6.70	(14)	6.79	(18)	13.54	(11)	16.08	(2)	18.71	(14)	1.00	(2)	4.54	(19)	
71	1140 1107	-	0.27	(10)	0.23	(12)	0.47	(20)	0.25	(8)	0.43	(10)	0.30	(6)	0.34	(5)	0.43	(15)	0.41	(4)	0.53	(9)	0.61	(2)	0.50	(5)	0.23	(1)	0.57	(6)
72	1152 1130	111620.47	(20)	0.92	(16)	0.85	(18)	1.08	(3)	1.59	(12)	-	-	0.26	(30)	0.24	(19)	0.27	(9)	1.22	(15)	1.94	(6)	1.03	(7)	-	-	-	-	
73	1174 1156	-	0.97	(18)	1.34	(17)	1.46	(5)	0.97	(18)	1.85	(10)	0.59	(10)	0.54	(8)	0.65	(4)	0.43	(7)	2.24	(25)	2.78	(3)	2.06	(15)	0.36	(10)	0.68	(6)
80	1284 1287	12910.61	(19)	0.95	12	0.67	(3)	0.47	(9)	0.73	(12)	0.26	(3)	0.28	(6)	0.58	(4)	0.32	(14)	1.06	(16)	0.81	(15)	0.65	(14)	0.40	(6)	0.60	(12)	
83	1320 1292	12977.66	(9)	8.90	(7)	14.55	(5)	10.77	(2)	16.94	(7)	1.42	(14)	2.90	(7)	1.16	(11)	5.52	(8)	15.81	(17)	28.34	(8)	16.09	(16)	0.29	(19)	3.27	(3)	
88	1345 1340	1329.20.80	(11)	38.58	(5)	62.53	(7)	62.57	(6)	69.31	(6)	4.12	(5)	8.77	(2)	24.14	(12)	32.01	(8)	86.41	(15)	95.26	(3)	80.70	(13)	0.91	(17)	9.50	(8)	
89	1366	-	-	0.21	23	0.30	(9)	-	0.31	(20)	0.20	(11)	0.16	(22)	0.20	(16)	0.16	(11)	0.48	(13)	0.31	(15)	0.37	(11)	0.22	(13)	0.21	(8)		

Table 2. Continued

Peak number ¹ _k (s) ² _k (s)	Chemical groups	sweet						medium sweet																							
		Malvasia			Tinta Negra			Bual			Tinta Negra																				
		SY B ^c	10Y B	15Y B	18Y ^d	20Y	3Y B	5Y B	10Y B	15Y B	17Y	19Y	3Y B	5Y B																	
84	4,5-Dimethyl-2-phenyl-1,3-dioxolane (m/z = 196, 152, 77, 43) ^f	R.I. 1320 -	1.44	(16)0.66	(13)0.49	(14)0.61	(2)	1.06	(6)	-	0.32	(13)0.97	(9)	0.74	(11)2.00	(9)	1.60	(4)	1.53	(13)0.39	(20)0.56	(5)									
93	1,1-Diethoxynonane	R.I. 1388	1401	1381	2.74	(4)	5.25	(17)4.49	(3)	4.15	(6)	14.28	(14)7.78	(1)	5.76	(4)	3.76	(4)	2.29	(9)	4.42	(7)	8.86	(16)3.52	(11)2.10	(14)4.14	(16)				
98	1,1-Diethoxydecane	R.I. 1485	1501	-	1.28	(14)2.63	(18)3.06	(9)	2.27	(2)	1.89	(6)	5.94	(11)1.81	(11)3.60	(18)2.63	(13)2.36	(10)5.32	(16)1.58	(16)1.21	(8)	2.31	(3)								
	Subtotal				439.28	(4)	637.08	(6)	641.90	(3)	869.95	(5)	963.25	(5)	423.83	(7)	432.30	(2)	583.86	(2)	696.06	(6)	949.17	(8)	982.46	(1)	1082.65	(4)	360.35	(10)605.91	(7)

^a Retention times in seconds (s) for first (¹t_R) and second (²t_R) dimensions. ^b RI: retention index obtained through the modulated chromatogram. ^c RI: retention index reported in the literature for one dimensional GC with a 5%-phenyl-methylpolysiloxane GC column or equivalent. ^d RI: retention index reported in the literature for a comprehensive GC × GC system with Equity-5 for the first dimension. ^e Y: blend wine; years, average age. ^f Y: vintage wines; age expressed in years. ^g Mean of three replicates. ^h Relative standard deviation, expressed in percentage. ⁱ Tentatively identified based on mass spectra.

(Figure 2). According to ²D (polarity), it was observed that furan, tetrahydrofuran, alkyl furans (2-ethoxytetrahydrofuran and 2-pentylfuran), and furanic acetal (2-diethoxymethylfuran) exhibited similar polarity. The polarity of these furans increase by the presence of an aromatic ring (–C₆H₅) (e.g., benzofuran) or a formyl (–CHO) (e.g., furanic aldehydes) group. Concerning furanic aldehydes, the presence of a second –CHO group in the α'-position related to the heteroatom of the heterocyclic ring, e.g., 5-formylfurfural (¹t_R = 432 s, ²t_R = 3.94 s) results in a considerable increase in polarity compared to the remaining furanic aldehydes, which were located in the chromatographic space of ¹t_R = 198–564 s and ²t_R = 1.10–1.90 s (Figure 2). However, furanic alcohols showed a high polarity compared to furanic aldehydes; consequently, an increase was observed in ²D (²t_R = 2.64–3.75). Although, Furaneol (¹t_R = 444 s, ²t_R = 1.33 s) is an exception, resultant from the presence of two methyl (–CH₃) groups in α- and α'-positions related to the heteroatom, a decreasing in polarity was observed. According to ²D, the elution order of furans under study based on their functional groups was alkyl ~ acetal < aromatic ring (benzofuran) < aldehyde < alcohol.

The lactones were organized in lactone/alkyl lactone, anhydride, enolic lactone, and aromatic lactone (Figure 3). The lactone/alkyl lactones are the less polar type; thus, lower ²D values (²t_R = 0.87–3.30 s) were observed. γ-Crotonolactone (¹t_R = 282 s, ²t_R = 4.15 s) is an exception; the high ²t_R was explained due to the presence of π-bound in the structure and the absence of the –CH₃ group.⁵⁶ The anhydride showed higher polarity (²t_R = 4.38–4.90 s) when compared to lactone/alkyl lactones due to the presence of RC=OR' group in the α'-position instead of H. Nevertheless, two exceptions were observed, namely, 3-methylenedihydro-2,5-furandione (¹t_R = 306 s, ²t_R = 2.99 s) and 3,4-dimethyl-2,5-furandione (¹t_R = 396 s, ²t_R = 2.02 s), which demonstrated lower polarity compared to others, due to the a weak π-bound in the β-position, and the two –CH₃ groups in β- and β'-positions of heterocyclic ring, respectively. Pantolactone (¹t_R = 396 s, ²t_R = 5.90 s), the unique enolic lactone detected, showed the highest polarity of lactones, which may be explained by the presence of the –OH group in the β-position of the heterocyclic ring. Thus, according to ²D, the elution order of lactones under study based on their functional groups was alkyl < aromatic ring < ketone (anhydride) < alcohol.

The volatile phenols were organized in phenolic ether, alkyloxy phenol, phenol/alkyl phenol, phenolic aldehyde, phenolic ester, and phenoxy alcohol (Figure 4). According to ²D, the phenolic ether exhibited the lowest polarity (²t_R = 0.92–1.21 s) compared to alkyloxy phenol, phenol/alkyl phenol, phenolic aldehyde, phenolic ester, and phenoxy alcohol. The presence of the –OH group in these former compounds increases their polarity; consequently, higher ²t_R values were observed. Intermediate polarity was observed for alkyloxy phenol (²t_R = 1.64–2.50 s), followed by phenolic ester (e.g., ethyl vanillate, ²t_R = 2.74 s), phenoxy alcohol (e.g., 2-phenoxyethanol, ²t_R = 3.18 s), and phenolic aldehyde (e.g., vanillin, ²t_R = 4.71 s). The alkyl phenols, namely, *o*-cresol, *p*-cresol, and *p*-ethylphenol showed the highest polarity (²t_R = 4.62–5.78 s), which may be explained by the absence of an –OCH₃ group in the α-position compared to all other phenols studied. Thus, according to ²D, the elution order of volatile phenols under study based on their functional groups was ether (phenolic ether < alkyloxy) < ester < aldehyde < alcohol (alkyl phenol).

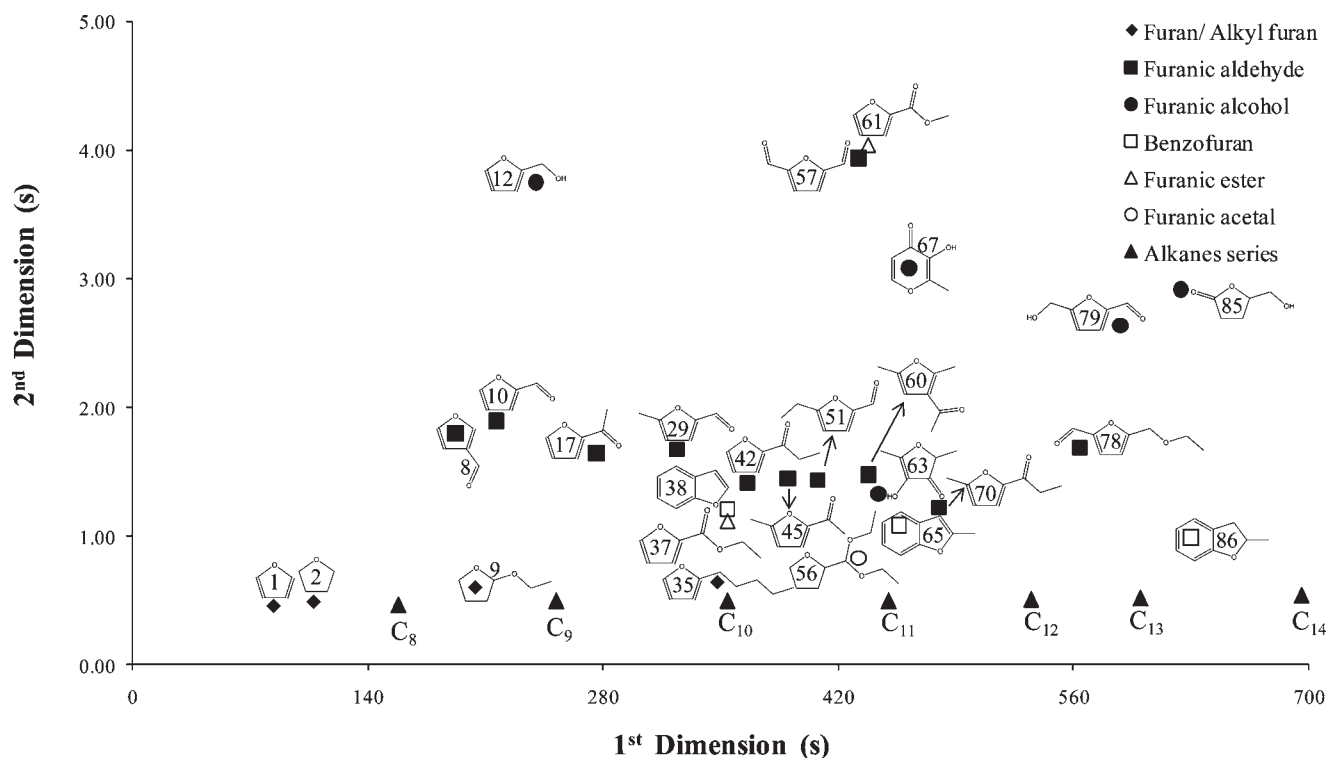


Figure 2. Peak apex plots of furans identified in Madeira wine (attribution of the peak number is shown in Tables 1 and 2).

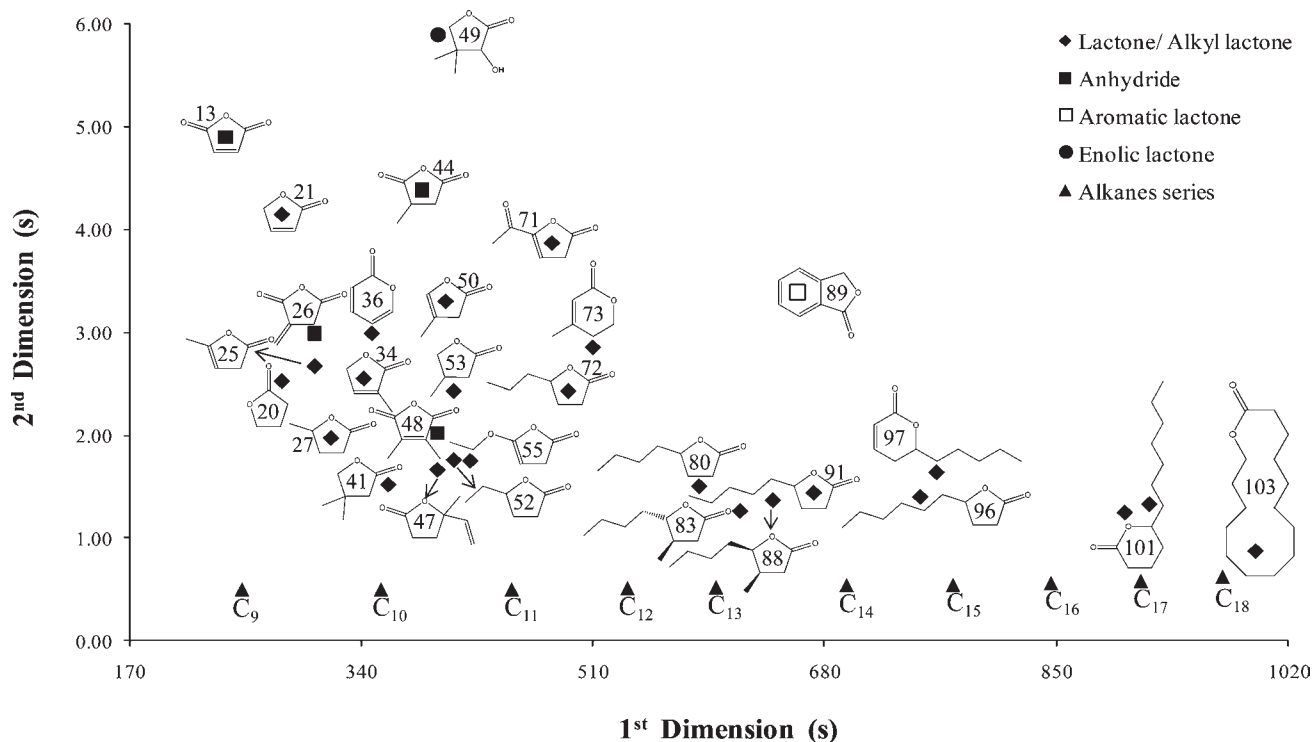


Figure 3. Peak apex plots of lactones identified in Madeira wine (attribution of the peak number is shown in Tables 1 and 2).

Concerning the acetals, three types of structures were observed: alkyl/heterocyclic acetal, alkyl/heterocyclic acetal alcohol, and aromatic acetal. Previous studies only allowed the detection of heterocyclic acetal alcohols.^{4,10,11,20} The alkyl/heterocyclic acetals are the less polar compounds; afterward,

they are the first compounds eluted according ²D. For these compounds, the maximum ²t_R achieved was 0.82 s. 2-Diethoxyethanol (alkyl acetal alcohol, ¹t_R = 294 s, ²t_R = 1.26 s) and 4,5-dimethyl-2-phenyl-1,3-dioxolane (aromatic acetal, ¹t_R = 618 s, ²t_R = 1.07 s) showed an intermediate polarity due to the presence

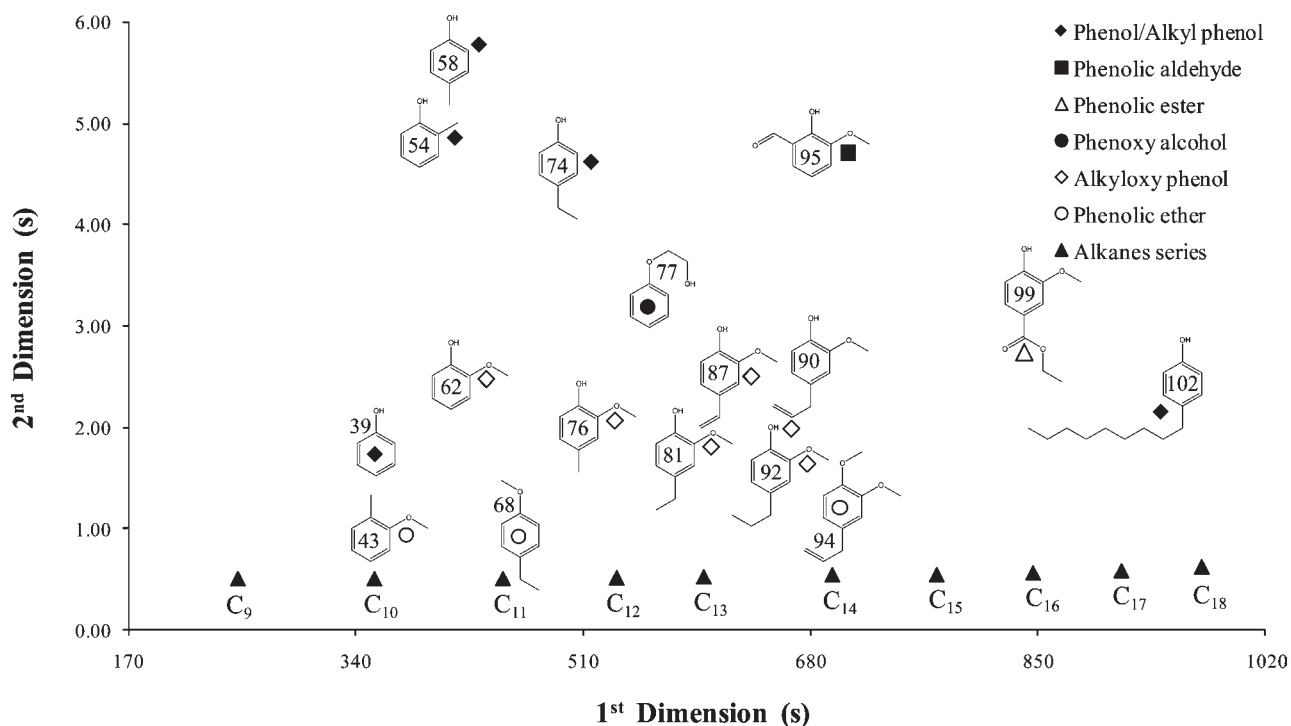


Figure 4. Peak apex plots of volatile phenols identified in Madeira wine (attribution of the peak number is shown in Tables 1 and 2).

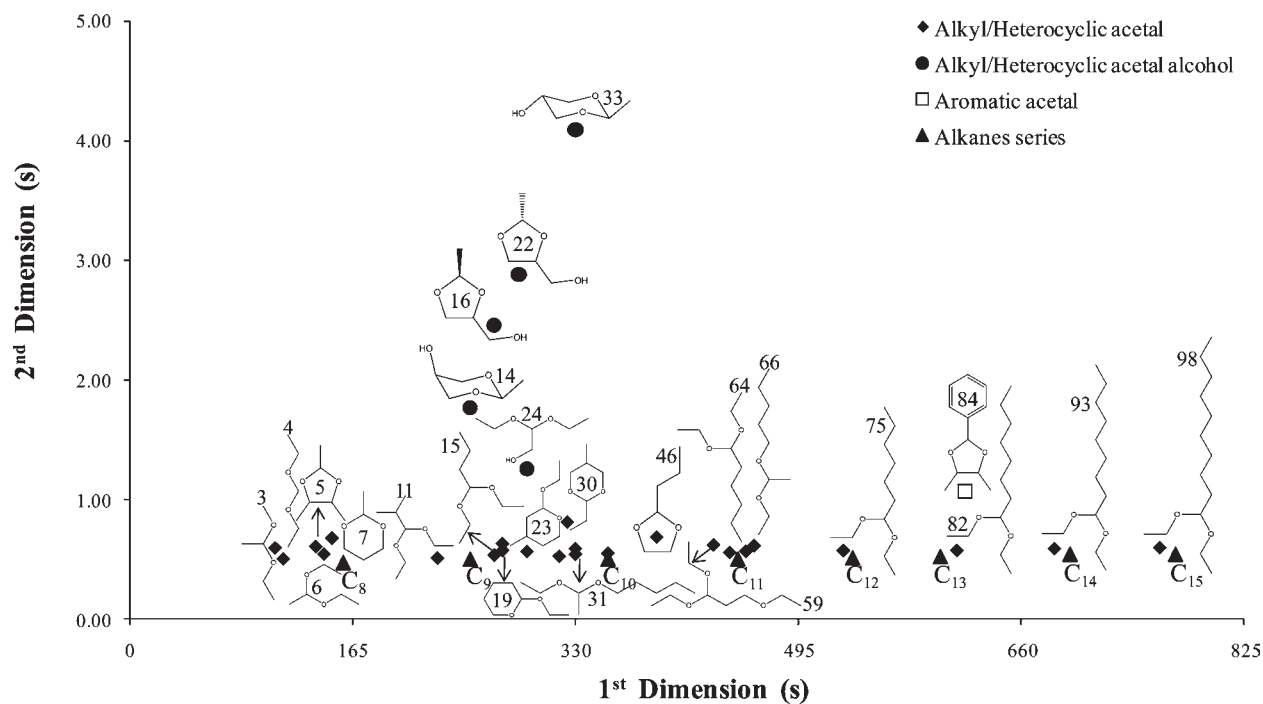


Figure 5. Peak apex plots of acetals identified in Madeira wine (attribution of the peak number shown is in Tables 1 and 2).

of the $-OH$ and $-C_6H_5$ groups. Otherwise, the four heterocyclic acetal alcohols located in the chromatographic space of $^1t_R = 252-330$ s and $^2t_R = 1.77-4.09$ s showed the highest polarity compared to heterocyclic acetals due to the $-OH$ group in their structure. Thus, according to 2D , the elution order of acetals under study was alkyl < aromatic ring < alcohol. Regarding the peak apex of the acetals displayed in Figure 5, it is possible

to observe the advantage of the second dimension for the analysis of different acetals.

The structured 2D chromatographic profile arising from 1D volatility and 2D polarity was observed within each chemical group based on the properties and positions of their functional groups, which allow more reliable identifications. Globally, based on the functional group of the chemical families under study, the

2t_R values increase in this way: alkyl < ether < ester < aromatic ring < ketones ~ aldehydes < alcohol. This information is especially useful for classifying unidentified compounds.

Establishment of Potential Age Markers. The GC peak area and RSD (relative standard deviation) values of furans, lactones, volatile phenols, and acetals obtained using HS-SPME/GC \times GC-ToFMS methodology are listed in Table 1 for dry and medium dry, and in Table 2 for sweet and medium sweet Madeira wines.

Furans. The furanic aldehydes, such as 2-furfural and 5-methyl-2-furfural, were the predominant furans tentatively identified in the Madeira wines under study. Other furans were also tentatively identified, namely, 2-ethoxytetrahydrofuran, 2-acetylfuran, ethyl 2-furoate, 5-formylfurfural, 5-ethoxymethyl-2-furfural, 5-hydroxymethylfurfural, and 5-hydroxymethyl-dihydrofuran-2-one. Some of these furans have been reported as age markers of Madeira wine, namely, ethyl 2-furoate, 5-ethoxymethyl-2-furfural, and 5-hydroxymethylfurfural, and a similar trend during aging was observed.^{4,12} From the total of 26 furans tentatively identified, only 8 compounds were previously identified in Madeira wines (see Table 1), and as far we know, the 18 furans listed in Tables 1 and 2 are detected for the first time in Madeira wine. In order to evaluate the trend of furans, lactones, volatile phenols, and acetals with the aging process, correlations between GC peak area and age were computed. This approach was only applied to Malvasia and Bual Madeira wines, as they are the unique varieties under study that presented samples with 5 different ages. The other varieties under study only have wines with 2 different ages. A correlation ($r = 0.87$) between total GC peak area of furans and age was found for Bual, while a lower correlation ($r = 0.56$) was achieved for Malvasia, which means that, for Bual, the total GC peak area of furans was 76% related with age (expressed as r^2 (coefficient of determination)), whereas for Malvasia, it was only 31%. Furthermore, from 26 furans tentatively identified, only 2 are the highest and positively correlated with wine age, namely, 3-furfural ($r = 0.88$ and 0.78 , respectively for Malvasia and Bual), and ethyl 2-furoate ($r = 0.96$ and 0.99). Thus, these compounds may be suggested as potential age markers for Malvasia and Bual varieties.

Lactones. γ -Butyrolactone and *cis*-oak-lactone were the main lactones detected. Regarding the two isomers of whisky lactones (*cis*- and *trans*-oak-lactones), the *cis* isomer showed the highest GC peak area than *trans*-isomer. As previously reported, in the wine acidic medium, an easier extraction of *cis*-oak-lactones occurs.¹⁶ γ -Lactones are among the most important compounds from a sensory point of view, and their content tends to increase during the aging process in oak casks (Tables 1 and 2). Similar trends of γ -lactones during aging were achieved for Madeira and red wines by Câmara et al.⁴ and Cérdan et al.,⁵ respectively. From the total of 30 lactones tentatively identified, only 9 compounds were previously identified in Madeira wines (see Table 1), and as far we know, the 21 lactones listed in Tables 1 and 2 are detected for the first time in Madeira wine. The total GC peak area of lactones showed a high correlation ($r = 0.93$) with age in Malvasia, whereas for Bual, a slightly lower correlation ($r = 0.71$) was observed. Thus, for Malvasia the total GC peak area of lactones was 87% related with age, whereas for Bual, it was only 50%. Moreover, from the 30 lactones tentatively identified, only 5 are the highest and positively correlated with age for Malvasia and Bual varieties, namely, pantolactone ($r = 0.78$ and 0.77 , respectively, for Malvasia and Bual), γ -ethoxybutyrolactone ($r = 0.76$ and 0.92), γ -heptalactone ($r = 0.87$ and 0.72), *trans*-oak-lactone ($r = 0.83$ and 0.77), and *cis*-oak-lactone ($r = 0.93$ and 0.82).

Volatile Phenols. The predominant volatile phenols detected were *o*-guaiacol, *p*-ethylphenol, and *p*-ethylguaiacol. During aging, for wines obtained from the Sercial grape variety (alcoholic degree of 16.8%⁴) these compounds increase more remarkably, whereas for Bual wines (17.8%⁴), a considerable decrease was observed. According to Dias et al.,⁵⁷ the content of *p*-ethylphenol and *p*-ethylguaiacol is more accentuated in wines with lower alcoholic degree because the high content reduces the microbial activity of yeast, making the synthesis of ethylphenols difficult. From the total of 17 volatile phenols tentatively identified, 11 compounds were previously identified in Madeira wines (see Table 1), and as far we know, the 6 volatile phenols listed in Tables 1 and 2 are detected for the first time in Madeira wine. Conversely, volatile phenols did not show a linear trend with age for Malvasia ($r = 0.05$) and Bual ($r = 0.05$) varieties. In addition, the individual analysis of all volatile phenols revealed that the GC peak area of these components were not correlated with wine age, with the exception of *p*-cresol ($r = 0.79$ and 0.75 , respectively for Malvasia and Bual). This behavior may be explained as the volatile phenols present several origins, i.e., oak, microbiological activity (e.g., *Brettanomyces* and *Dekkara*), and hydroxycinnamic acids of wine,²³ among others.

Acetals. The major acetals detected were diethoxymethane, 1,1-diethoxyethane, 1,1-diethoxy-2-methyl-propane, 1,1-diethoxy-3-methylbutane, and 1-(1-ethoxyethoxy)-pentane. From the total of 30 acetals tentatively identified, only 4 compounds were previously identified in Madeira wines (see Table 1), and as far we know, the 26 acetals listed in Tables 1 and 2 are detected for the first time in Madeira wine. The white varieties (Malvasia, Bual, Sercial, and Verdelho) presented high total GC peak area of acetals than the red one (Tinta Negra). Similar results were achieved by Cutzach et al.,⁵⁸ as the high content of polyphenols in red varieties slowed the oxidation reaction and combined easily with acetaldehyde. For acetals, high correlation with age was observed for Malvasia ($r = 0.95$), and Bual ($r = 0.95$). Similar correlation was observed by Câmara et al.^{4,10} for heterocyclic acetals alcohols previously identified in Madeira wines. Additionally, other acetals such as 1,1-diethoxy-2-methyl-propane ($r = 0.81$ and 0.86 , respectively, for Malvasia and Bual), 2,2-diethoxyethanol ($r = 0.96$ and 0.84), 1-(1-ethoxyethoxy)-pentane ($r = 0.96$ and 0.81), 1,1-diethoxypentane ($r = 0.81$ and 0.94), 2-propyl-1,3-dioxolane ($r = 0.86$ and 0.87), and 1,1-diethoxyhexane ($r = 0.95$ and 0.83) showed high correlation with age for Malvasia and Bual varieties.

In sum, total GC peak area of acetals showed the highest and positive relationship ($r \geq 0.95$) with age, whereas the volatile phenols showed the lowest one ($r \leq 0.05$). Furthermore, from the 103 volatiles tentatively identified to which belong furans, lactones, volatile phenols and acetals, 3-furfural, ethyl 2-furoate, pantolactone, γ -ethoxybutyrolactone, γ -heptalactone, *trans*-oak-lactone, and *cis*-oak-lactone, *p*-cresol, *trans*-dioxane, *trans*-dioxolane, 1,1-diethoxy-2-methyl-propane, 2,2-diethoxyethanol, 1-(1-ethoxyethoxy)-pentane, 1,1-diethoxypentane, 2-propyl-1,3-dioxolane, and 1,1-diethoxyhexane exhibited the highest and positive correlations ($0.71 < r < 0.99$) with wine age. Thus, these compounds may be suggested as potential age markers of Malvasia and Bual wines. In a second step, PCA was applied to autoscaled GC peak areas of furans, lactones, volatile phenols, and acetals, in order to extend the study of potential age markers to all noble varieties (Malvasia, Bual, Sercial, and Verdelho) and to the major variety (Tinta Negra) used to produce Madeira wine, which includes different types of wines and ages as described below.

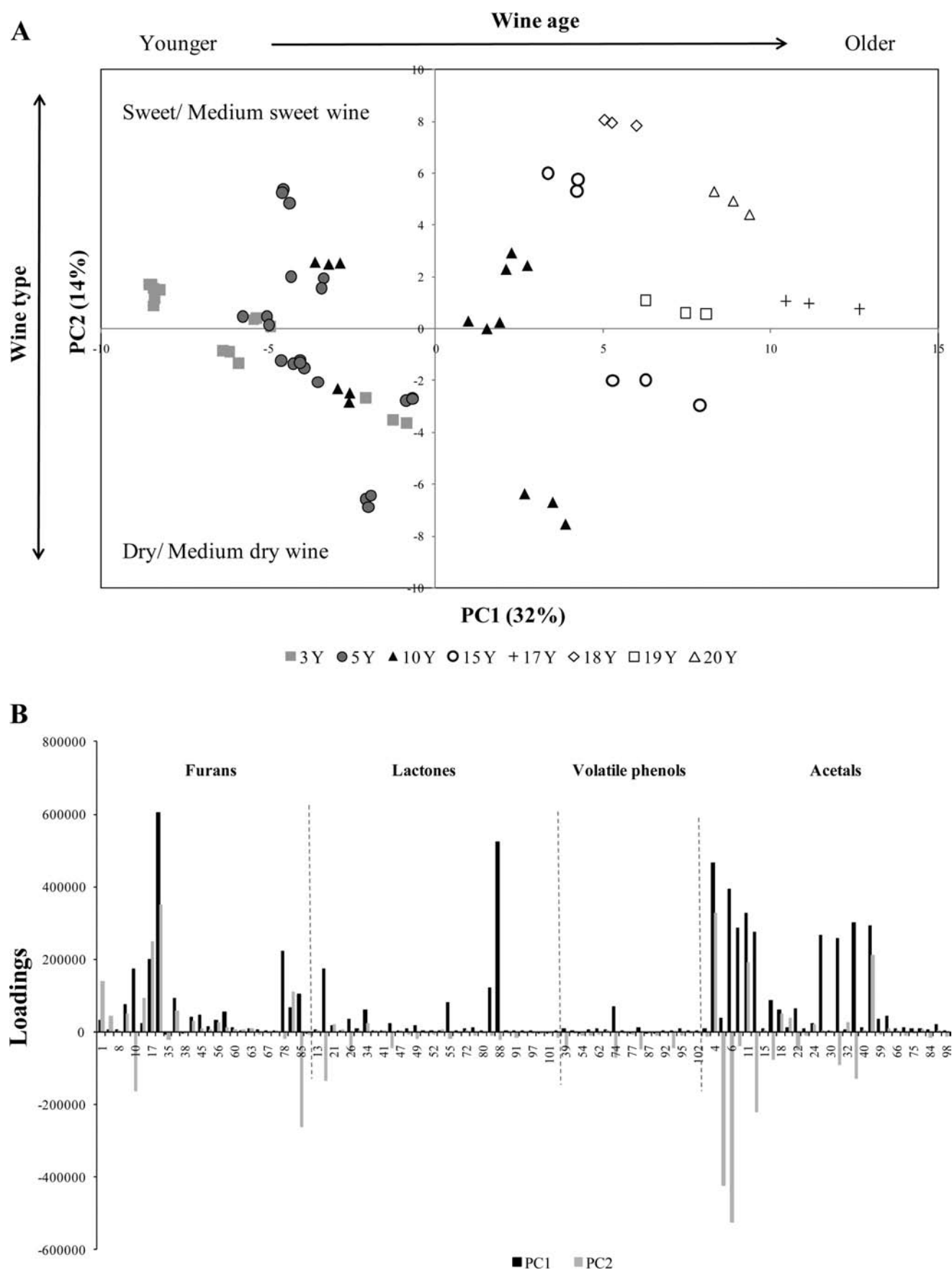


Figure 6. PC1 \times PC2 scores scatter plot (A) and loadings plot profile (B) of GC peak area (attribution of the peak number is shown in Tables 1 and 2).

Principal Component Analysis. GC peak area of 103 analytical variables (volatile compounds) of the 23 Madeira wines were submitted to a PCA procedure, in order to search for the main sources of variability, to characterize the samples as a function of the detected compounds, and to establish a possible relationship among furans, lactones, volatile phenols and acetals, and wine age. Figure 6A shows the score scatter plot of the two first principal components (which explains 46% of the total variability of the data set), allowing us to organize the Madeira wines by age (as a function of PC1 axis). Figure 6B represents the corresponding loadings plot profile which establishes the relative importance of each volatile for the observed sample distribution of Figure 6A. PC1, which explains 32% of the total variability, allow one to distinguish Madeira wines as a function of their age. PC2, explaining 14% of the total variability, shows the organization of the Madeira wines according to their type. With the exception of Tinta Negra, which may be used to produce all types of wines, all other varieties were used to produce a specific type of wine. Sweet (Malvasia) and medium sweet (Bual) Madeira wines (PC2 positive) are characterized by 5-methyl-2-furfural and diethoxymethane, whereas dry (Sercial) and medium dry (Verdelho) were placed in PC2 negative and are described by 1,1-diethoxyethane and 2,4,5-trimethyl-1,3-dioxolane. As can be observed in Figure 6A, the Madeira wines could be organized by their type, as well as grape variety according to PC2. According to PC1, the Madeira wines with 3 and 5Y are projected in PC1 negative, whereas 10Y is near the origin, except for two Madeira wines, which were located near of 5Y, as these wines correspond to the averaged aging period (blended wines). The Madeira wines with 15, 17, 18, 19, and 20Y are placed in PC1 positive. Moreover, the Madeira wines could be organized by their style (vintage or blends) according to PC1. The blend Madeira wine is projected in PC1 negative and is near the origin, whereas the vintage is placed in PC1 positive. Taking into account the loadings plot (Figure 6B), the chemical compounds used as potential age markers were predominantly acetals, namely, diethoxymethane, 1,1-diethoxyethane, 1,1-diethoxy-2-methylpropane, 1-(1-ethoxyethoxy)-pentane, *trans*-dioxane, and 2-propyl-1,3-dioxolane, and from the other chemical groups studied, 5-methylfurfural (furan) and *cis*-oak-lactone showed similar contribution to acetals. Finally, it is important to point out that, from these 8 compounds, *cis*-oak-lactone, *trans*-dioxane, 1,1-diethoxy-2-methylpropane, 1-(1-ethoxyethoxy)-pentane, and 2-propyl-1,3-dioxolane were previously (in Establishment of Potential Age Markers) proposed as potential age markers for the Malvasia and Bual varieties. These results suggest that among the chemical groups under study, the acetals are the most important group that could be used as potential age markers of Madeira wines, independently of the variety and the type of wine. Despite the fact that acetals are formed during the fermentation step, its content increases remarkably during the aging process, which may be explained by the oxidative condition that occurred during this step. These oxidative conditions contribute to the increase of aldehyde content, mainly acetaldehyde, and acetals.¹⁰ Moreover, GC × GC-ToFMS offered a very useful approach to identify these chemical groups due to the orthogonal systems that reduce coelution and improve the quality of the selection of volatile compounds under study, contributing to the establishment of new potential Madeira wine age markers.

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